

**Description**

Power Taq 2X Master Mix with Blue Dye is a Ready to Use, High fidelity mix ,containing DNA polymerase, dNTPs, MgCl<sub>2</sub>, KCl and other stabilizers. This product is suitable for conventional PCR amplification from gDNA from Plants, Animals, Human. The template can be purified DNA, bacterial colonies/bacteria liquid, crude extract or cDNA, etc. This product can use complex **genomic DNA** as a template to amplify a target fragment of 5 kb in length or a simple template such as **lambda DNA** to amplify a target fragment of 10 kb in length. It is suitable for applications such as PCR reaction, colony PCR identification, Plant Transgene identification, gene cloning, vector construction.

**Kit Contents**

Contents	FS-T-91702-10	FS-T-91702-25
Power TAQ 2X Master Mix with blue Dye	10 ml	25 ml

1 ml= 40 reactions (Volume of 50 µl)

**Applications:** Conventional PCR, High Fidelity, GC-content, High speed

**Source:** The DNA Polymerase gene was induced and expressed in E.coli and obtained by separation and purification.

**Thermal inactivation:** No  
**5'-3' exonuclease activity:** No  
**3'-5' exonuclease activity:** Yes

**Storage Conditions**

Store all components at -20°C in a non-frost-free freezer.

**Quality Control**

No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

**Recommended Protocol**

This standard protocol applies to a single reaction where only template, primers, and water need to be added to the Power TAQ 2X Master Mix with Blue Dye. For multiple reactions, scale-up volume of reaction components proportionally. All reagents should be thawed on ice, gently mixed and briefly centrifuged before use.

1. Thaw reagents at room temperature. Mix thoroughly and then place on ice immediately after thawing.
2. Assemble reaction tubes on ice whenever possible to avoid premature, nonspecific polymerase activity.

**Reaction Conditions**

Component	25 µl	50 µl	Final
Power TAQ 2X Master Mix With Blue Dye	12,5 µl	25 µL	1X
10µM Forward Primer	0.5 µL	1 µL	0.2 µM
10µM Reverse Primer	0.5 µl	1 µL	0.2 µM
Template DNA*	Variable	Variable	<300 ng
Water, RNase-Free	up to 25 µl	up to 50 µl	N/A

\*High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed below

**DNA TEMPLATE**

DNA	Input Amount
Plants, animals and human gDNA	10 ng~100 ng
E.coli , lambda gDNA	500 pg-200 ng
Plasmid DNA	1 pg~10 ng

**Note:** If the DNA template is obtained from a cDNA synthesis reaction, the template volume should be less than 10% of the total reaction volume. If long fragments are amplified, the amount of template input should be increased appropriately

**Recommended PCR Program**

Step	Temp	Time	Cycles
Pre-denaturation	98°C	45 s	1
Denaturation	98°C	10 s	30
Annealing	55-65°C	30 s	
Extension	72°C	20-30s/kb	
Post-extension	72°C	5min	1
Hold	4-12°C	∞	1

**2. Primers :**

Oligonucleotide primers are typically 20-40 nucleotides in length with a **GC content of 40-60%**. Primers can be designed and analyzed using software such as Primer 3 The final concentration of each primer in the PCR reaction system should be in the range of 0.1-1 µM.

**3. Denaturation:**

98°C pre-denaturation for 45 s can fully denature most DNA templates. In the case of high complexity DNA templates, the pre-denaturation time should be extended up to 3 minutes for fully denaturation.

Generally, the recommended denaturation condition for low-complexity DNA templates is 98°C, 5-10 s

**4. Annealing:**

The annealing temperature of Power TAQ 2x Master Mix with Blue Dye is usually higher than other PCR polymerases.

1. Generally, primers longer than 20 nt are annealed at (lower primer T<sub>m</sub>+3)°C for 10-30 s;
2. When the primers are shorter than 20 nt, an annealing temperature equivalent to the lower primer T<sub>m</sub>. When using a new primer set for PCR reaction, we recommend a gradient PCR to determine the optimal annealing temperature. In a two-step amplification protocol, the annealing temperature should be set to the extension temperature.

**5. Extension:**

The recommended extension temperature is 72 °C . The extension time depends on the length and complexity of the amplicon. For the low-complexity amplicons (plasmid DNA), the extension condition is 10 s/kb. For high-complexity amplicons, it is recommended to increase the extension time to 20-30 s/kb. In some cases, the extension time for cDNA templates should be less than 1 min/kb

**6. Cycles:**

To obtain enough yield of PCR products, 25-35 cycles are recommended.